

B-71 FATE OF URINARY COMPONENTS AND PHARMACEUTICALS IN STORAGE PROCESS OF URINE

○ Takashi KAKIMOTO^{1*}, Shinya HOTTA², Hitoshi SHIBUYA³, Haruki SUZUKI³
and Naoyuki FUNAMIZU²

¹Center for environmental science in Saitama.

(Kamitanadare 914, Kazo-shi, Saitama 347-0115, Japan)

²Department of Environmental Engineering, Graduate School of Engineering, Hokkaido University.

(Kita-13, Nishi-8, Kita-ku, Sapporo, 060-8628, Japan)

³Division of Laboratory and Transfusion Medicine, Hokkaido University Hospital.

(Kita-14, Nishi-5, Kita-ku, Sapporo, 060-8648, Japan)

* E-mail: kakimoto.takashi@pref.saitama.lg.jp

1. INTRODUCTION

Separation of urine at source has been a basic idea for more sustainable management of wastewater in household¹⁻⁷⁾. It is possible to prepare two main processes for source-separated urine: first one is onsite storage and second one is treatment of micro pollutants and recovery of nutrients *etc.* Various kinds of treatment and recovery methods have been investigated, however, compared to interests in phosphate, nitrogen and micro-pollutants, discussion about organic matter seemed to be not enough especially on the change in the character of urine during storage process. Therefore objectives of this study were 1) to prepare the list of urine components mainly focused on organic matter 2) to monitor the evolution of the urine components in storage process.

2. MATERIAL AND METHODS

(1) Collection and storage test of fresh urine

Fresh urine was collected from 189 volunteers (145 males and 44 females) in Hokkaido University within 4 hours around noon. After the collection, pharmaceuticals of carbamazepine, diclofenac and metoprolol were immediately added to fresh urine for pharmaceuticals' monitoring. For organic matter monitoring, 100 ml of the fresh urine was put into 100 ml of glass bottles (storage units). Every storage unit was stored in incubator (30 °C).

(2) Pretreatment and DOC determination

In 90 days storage test, sampling and analysis were conducted for several times. In every sampling

process, urine pH was measured immediately after the storage units were opened. Then urine was filtrated by membrane filter with pore size of 0.45μm and UF membrane (MWCO 10 kDa, 1 kDa) and dialysis tube (MWCO 100 Da). By using these membrane, organic matter in urine was fractionated and their DOC were determined by TOC analyzer.

(3) Determination of amino acid, organic acid and pharmaceuticals in urine

Determination of organic acid was conducted by using the ion exclusion chromatography system. Amino acid analysis was conducted by using amino acid analyzer (HPLC: Modell-8500, Hitachi). For the determination of pharmaceuticals, clean up and determination methods were followed by the OASIS technical notebook.

3. RESULTS AND DISCUSSIONS

(1) Compounds in very fresh human urine

Table 1 lists the comparison of major components measured in this study. As seen, most comparable values obtained in this study were similar to previous reports^{8,9)}. From the data obtained in this study, it was found that approximately 84% of organic matter in fresh urine was still unknown even though various compounds were determined; nitrogen species in urine were mostly described by four nitrogen species: urea, ammonia, uric acid and creatinin; major species of organic acid from fresh urine were the acetic acid, the lactic acid and the propionic acid; in amino acids the taurine and the histidine were slightly higher than other species.

(2) Fate of the fresh urine for 90 days storage

a) Transformation of organic matter

In this study, we monitored the organic matter in urine during the storage process. Figure 1 shows the evolution of DOC including urea and DOC except urea. As seen, DOC except urea was on the stable level throughout the storage process whereas the DOC including urea constantly decreased. this indicated that main reduction of organic matter was degradation of urea. Regarding organic matter except urea, 10% reduction was found. Since stored urine usually provide pH more than 7 with ammonia on high level, then it might be difficult to assume methane production as anaerobic reaction of organic matter^{10,11)}. Udert *et al.*¹²⁾ reported more than 80% of dissolved COD of urine was biodegradable in aerobic condition whereas they discussed possibility for anaerobic biodegradation of organic matter in stored urine by citing previous reports^{13,14)}.

Figure 2 shows the molecular weight distribution of the DOC except urea in storage process of urine. Simple result was found: 74% of organic matter were distributed in 100 Da - 1 kDa fraction in fresh urine whereas 84% of organic matter in stored urine presented in the fraction of less than 100 Da. It was found that the most of organic acid increased in the first 15 days during storage process (data not shown). That also supported the transformation of organic matter into smaller size.

c) Fate of pharmaceuticals

As shown in Fig.3, significant decrease of Tetracycline was found in stored urine at 30°C. Kakimoto and Funamizu¹⁵⁾ showed that the decomposition of Tetracycline profoundly related to levels of phosphate ion and Ammonia level in composting process of human feces. In urine, concentrations of phosphate and ammonium were much higher than in usual wastewater. It was therefore seemed to be reasonable that the tetracycline could decompose in storage process of urine. Slight decrease of the metoprolol was observed. Other pharmaceuticals used in this study did not decrease remarkably. It might be possible to assume that further three chemicals (Carbamazepine, Levofloxacin and Diclofenac) could be stable in simple storage process of urine. Carbamazepine was very stable even in the electro oxidation process¹⁶⁾. Several results from previous studies showed that Levofloxacin was hard to decompose in wastewater treatment plant¹⁷⁾. Diclofenac could decompose in wastewater treatment plant mainly because of bacteria³³⁾. Biodegradability of the diclofenac does increase in

Table 1 Comparison of components of fresh urine

		Urine [1] ⁹⁾	Urine [2] ⁸⁾	Our results
Dilution	(-)	1	1	1
pH	(-)	6.2	7.2	6.5
DOC	mg-C/l	-	5546*	7300
COD	mg-C/l	-	3056	4950*
	mg-N/			
Total N	l	8830	-	6300
Urea	mg/l	-	12450	11800
	mg-N/			
Ammonia	l	463	254	300
		800-***		
Phosphate	mg-P/l	2000***	367**	1100**
Na+	mg/l	3450	2670	2940
Cl] <div style="display: inline-block; width: 1em; border-bottom: 1px solid black; position: relative; top: -2px;"> <div style="position: absolute; left: -2px; top: -2px;">]</div> </div>	mg/l	4970	3810	5180
Ca2+	mg/l	233	129	120
Mg ²⁺	mg/l	119	77	60
K+	mg/l	2737	2710	1760

*: Estimated by assumption of "DOC = C in Urea + C in COD".

**: Including all system species of phosphate ion

***: Total phosphorous

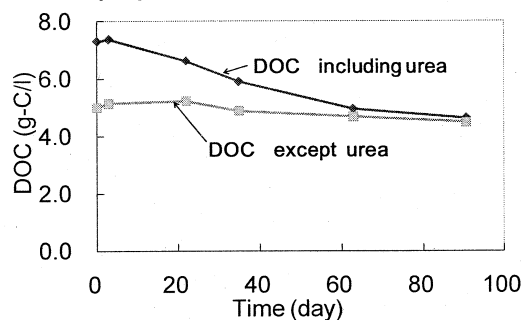


Figure 1 evolution of DOC in storage process

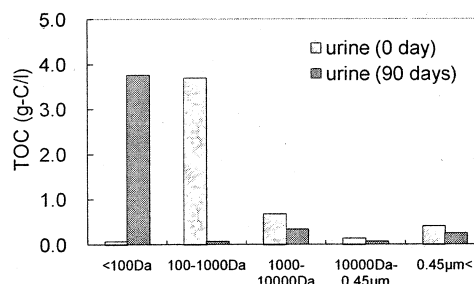


Figure 2 Molecular weight distribution of urine

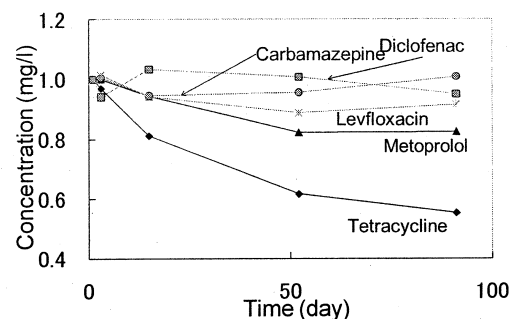


Figure 5 Pharmaceuticals in urine during storage process

acidic condition that provides non-dissociation type. However, pH condition was relatively higher than pH 9 in stored urine. Thus it might be hard for diclofenac to decompose.

4. CONCLUSION

Study on source-separated urine was conducted. Conclusions obtained in this study were following:

- (a) Lists of the components of very fresh urine were obtained: 40 species of amino acid, 7 species of organic acid, 4 species of nitrogen and 7 species of inorganic ion. However, more than 80% of organic carbon except Urea was still not determined in fresh urine.
- (b) 90 days storage provided smaller molecules. The majority of organic matter in fresh urine was found in the fraction of 100Da-1000Da whereas it shifted into the fraction less than 100Da after long storage. However the main compounds less than 100Da was still unknown. In the storage unit, more than 90% of organic matter was not carbonated but they remained in dissolved type even after 90days storage at mild temperature. Some species of organic acid temporally increased but finally decreased.
- (c) Declines of the Tetracycline and Metoprolol were found in simple storage unit with sealed at 30°C. However pharmaceuticals of Carbamazepine, Levofloxacin and Diclofenac were being on stable level throughout the storage process.

ACKNOWLEDGMENT: We thank all of volunteers with contribution to urine collection and all of staff in the Hokkaido University Hospital for their help in urine analysis of organic nitrogen, inorganic ion species. Mr.Yusuke Ino provided great helps with us especially in filtration process of urine. This work has been supported by CREST of the JST (Japan Science and Technology Agency).

Reference

- 1) Hellstrom, D. and Karrman, E., 1997. Exergy analysis and nutrient flows of various sewerage systems. *Water Science and Technology*, 35(9), 135-144.
- 2) Hellstrom, D., 2003. Exergy analysis of nutrient recovery processes. *Water Science and Technology*, 48(1), 27-36.
- 3) Larsen, T. and Gujer, W., 1996a. Separate management of anthropogenic nutrient solutions (human urine). *Water Science and Technology*, 34(3-4), 87-94.
- 4) Larsen, T. and Gujer, W., 1996b. The concept of sustainable urban water management. *Water Science and Technology*, 35(9), 3-10.
- 5) Larsen, T. A., Peters, I., Alder, A., Eggen, R., Maurer, M., and Muncke, J., 2001. Re-engineering the toilet for sustainable wastewater management. *Environmental Science and Technology*, 35(9), 192A-197A.
- 6) Larsen, T., Lienert, J., Joss, A. and Siegrist, H., 2004. How to avoid pharmaceuticals in the aquatic environment. *Journal of Biotechnology*, 113, 295-304.
- 7) Vinneras, B. and Jonsson, H. 2002. The performance and potential of faecal separation and urine diversion to recycle plant nutrients in household wastewater. *Bioresource Technology*, 84(3), 275-282.
- 8) Udert, K. M., Larsen, T. A., Biebow, M. and Gujer, W., 2003a. Urea hydrolysis and precipitation dynamics in a urine-collecting system. *Water Research*, 37(11), 2571-2582.
- 9) Ciba-Geigy, 1977. *Wissenschaftliche Tabellen Geigy, Teilband Ko'rperflu' ssigkeiten* (Scientific Tables Geigy. Volume: Body Fluids), 8th ed. Basel. In German.
- 10) Calli, B., Mertoglu, B., Inanc, B. and Yenigun, O., 2005. Effects of high free ammonia concentrations on the performances of anaerobic bioreactors. *Process Biochemistry*, 40(3-4), 1285-1292.
- 11) Udert, K. M., Larsen, T. A., and Gujer, W., 2006. Fate of major compounds in source-separated urine. *Water Science & Technology*, 54(11-12), 413-420.
- 12) Udert, K. M., Fux, C., Munster, M., Larsen, T.A., Siegrist, H. and Gujer, W., 2003b. Nitrification and autotrophic denitrification of source-separated urine. *Water Science and Technology*, 48(1), 119-130.
- 13) Hoglund, C., Stenstrom, T. A., Jonsson, H. and Sundin, A., 1998. Evaluation of faecal contamination and microbial die-off in urine separating systems. *Water Science and Technology*, 38(6), 17-25.
- 14) Hoglund, C., Vinneras, B., Stenstrom, T. A. and Jonsson, H. 2000., Variation of chemical and microbial parameters in collection and storage tanks for source separated human urine. *Journal of Environmental Science and Health*, A35(8), 1463-1475.
- 15) Kakimoto, T. and Funamizu, N. 2006. Effect of phosphate, ammonia and pH on the degradation of antibiotics in the composting toilet, *Environmental Engineering Research*, 43, 429-435.
- 16) Osawa, T. 2007. Electro-oxidation of pharmaceuticals in urine. Master thesis in Hokkaido University. *In Japanese*.
- 17) Yasojima, M., Kobayashi, Y., Nakada, H., Komori, K., Suzuki, Y. and Tanakam, H. 2005. Behavior of human antibiotics in wastewater treatment plants. *Environmental Engineering Research*, 42, 357-368.
- 18) Urase, T. and Kikuta, T. 2005. Separate estimation of adsorption and degradation of pharmaceutical substances and estrogens in the activated sludge process *Water Research*, 39(7), 1289-1300.